



SURVEILLANCE REPORT

Influenza virus characterisation

Summary Europe, June 2013

Summary

Over the course of the 2012–13 season, A(H1N1)pdm09, A(H3N2) and B/Victoria- and B/Yamagata-lineage influenza viruses have co-circulated in ECDC-affiliated countries. The relative prevalences of each has varied between countries.

- Type A and type B viruses have been detected in similar proportions but with type A peaking and declining slightly before type B.
- A(H1N1)pdm09 viruses have been detected at approximately twice the level of A(H3N2) viruses.
- The vast majority of A(H1N1)pdm09 viruses have remained antigenically similar to the vaccine virus, A/California/07/2009, but continued to show genetic drift with an increasing prevalence of genetic group 6 viruses.
- The vast majority of A(H3N2) viruses have been antigenically and genetically similar to cell-propagated A/Victoria/361/2011, the prototype vaccine virus for the 2012–13 influenza season.
- Viruses of the B/Yamagata-lineage have predominated over those of the B/Victoria-lineage.
- B/Victoria-lineage viruses have remained antigenically similar to cell-propagated reference viruses of the B/Brisbane/60/2008 genetic clade.
- B/Yamagata-lineage viruses formed two antigenically distinguishable genetic clades: clade 3 represented by B/Wisconsin/1/2010 (the recommended vaccine component for the 2012–13 influenza season) and, in increasing numbers, clade 2 represented by B/Massachusetts/2/2012 (the recommended vaccine component for the 2013–14 influenza season).

Viruses from specimens collected between 1 January 2013 and 31 May 2013, spanning the peak of the 2012–13 season, were received from 25 countries in the EU/EEA region at the MRC National Institute for Medical Research, WHO Collaborating Centre for Reference and Research on Influenza. A summary of specimens received is shown in Table 1.

The overall proportions of influenza type A (59%) and type B (41%) viruses received have become more similar, reflecting the decreasing proportion of influenza A towards the end of the season at the same time as the numbers of influenza virus detections were also falling. For type A, H1N1pdm09 viruses were received in greater numbers than H3N2 viruses (ratio 2:1). Among influenza B receipts, viruses of the B/Yamagata and B/Victoria lineages were received at a ratio of 4:1.

This report was prepared by Rod Daniels, Vicki Gregory and John McCauley on behalf of the European Reference Laboratory Network for Human Influenza (ERLI-Net), under contract to the European Centre for Disease Prevention and Control (ECDC).

 $[\]ensuremath{\mathbb{C}}$ European Centre for Disease Prevention and Control, Stockholm, 2013.

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Table 1. Summary of clinical samples and isolates received from ECDC-affiliated countries, with collection dates since 1 January 2013

MONTH	TOTAL RECEIVED	Α	H1N1	pdm09	Н	3N2	В	B Victor	ria lineage	B Yamag	gata lineage
Country			Number	Number	Number	Number		Number	Number	Number	Number
			received	propagated ¹	received	propagated ²		received	propagated ¹	received	propagated ¹
JANUARY											
Belgium	19	2	7	5	1	1				9	6
Czech Republic	11		9	9	2	2					
Denmark	3		1	1	2	2					
Estonia	21	2	9	5	2	1	5			3	3
Finland	7		3	3	3	3		1	1		
France	1		1	1							
Germany	9		2	2	2	2		1	1	4	4
Greece	7	1	2	1	3	1				1	1
Ireland	6		47	47	2	2		1	1	3	3
Italy	32		17	17	5	5		3	3	7	7
Latvia	7		3	3	3	3				1	1
Luxembourg	13		8	7		4				5	4
Malta Netherlands	24		18	2 1	1	1				5 1	5 1
Norway	2 4		1	3						1	1
Portugal	22		11	in process	4	4		5	in process	2	in process
Romania	8	1	5	in process 5	4	4		5 1	in process 1	2	in process 2
Slovenia	8 18	1	5 4	5 1	5	3	1	5	4	2	2
Spain	22	1	10	10	6	6	'	5	-	6	6
Sweden	8		10	10	3	3				5	in process
Sweden United Kingdom	6				5	3 5		1	1	5	in process
onited Kingdom	0				5	5					
FEBRUARY											
Belgium	12		2	in process	2	in process	8				
Bulgaria	22	1	7	7	2	2	Ů			12	12
Czech Republic	4			•	-	-				4	4
Greece	2		2	2						-	-
Hungary	12		6	4				3	3	3	2
Iceland	1		-	-				-	-	1	1
Italy	22		11	11	1	1		1	1	9	9
Luxembourg	5		1	0	4	0			-		-
Portugal	10		4	in process	-	-		4	in process	2	in process
Romania	12		7	7						5	5
Slovakia	11		2	2	3	3				6	in process
Slovenia	15		6	6	4	4		1	1	4	3
Spain	10		9	7	1	1					
Sweden	8		-		4	4				4	in process
United Kingdom	8		3	3	2	2		1	1	2	2
MARCH											
MARCH	45				3		8				
Belgium	15		4	in process	3	in process	8				
Bulgaria	2		1	1						1 1	1 1
Czech Republic	1							4			
Hungary	11 3		1	1	1	4		1	1	9 2	3 2
Iceland	-		1	1		1					
Italy	2			I						1	1
Luxembourg Portugal	1 13		5	in process				3	in process	1 5	1 in process
Romania	5	1	1	1	1	1		5	11 0100000	3	3
Slovakia	13	1	4	4	3	3		2	in process	4	in process
Slovenia	3			-	Ĩ	U U		2	1	1	1 1
Spain	9	1	3	2	4	4		-	•	2	1
Sweden	1		1	in process		-				_	
APRIL		1									
Belgium	3				2	in process	1				
Hungary	4	1						2	2	1	1
Iceland	3	1	1	1	1	1				1	1
Portugal	2	1	1	in process						1	in process
Romania	2		1	1						1	1
Slovakia	1	1								1	in process
Slovenia	2		1	1						1	1
Spain	3		1	0	1	1				1	1
МАҮ											
Iceland	3		1	1						2	2
	506	7	202	139	88	72	23	38	22	148	105
25 Countries			39	9.9%	17	7.4%		7	.5%	2	9.2%
25 Countries				59%					41%		
	1										

1. Propagated to sufficient titre to perform HI assay (the totalled number does not include any from batches that are in process) 2. Propagated to sufficient titre to perform HI assay in presence of 20nM oseltamivir (the totalled number does not include any from batches that are in process)

Influenza A(H1N1)pdm09 virus analyses

The results of HI assays carried out on influenza A(H1N1)pdm09 viruses since the May report [1] are shown in Table 2. All but one test virus, A/Hungary/02/2013, showed good reactivity with post-infection ferret antisera raised against the panel of reference viruses, including antiserum raised against the vaccine virus, A/California/7/2009, with this antiserum recognising the test viruses shown at titres within fourfold of its recognition of the homologous virus. As described previously [2], antiserum raised against A/Christchurch/16/2010, a virus from a genetic group not seemingly in circulation at present (group 4), reacted less well than the other antisera with the test viruses; this ferret antiserum reacted with 14 of the 15 test viruses, with titres reduced eightfold or greater compared with the titre of the antiserum with the homologous virus.

Antisera raised against several of the reference viruses reacted poorly with A/Hungary/02/2013, showing eightfold or greater reductions in titre compared to the titres of the antisera with their homologous antigens; HA gene sequencing of this virus showed that it carried a N156D substitution in HA1. Amino acid substitution or polymorphism in the 153–157 region of HA1 can affect the antigenicity of the virus and commonly emerges during propagation of viruses in cell culture. A/Hungary/02/2013, but for its reactivity with antiserum raised against the vaccine virus (A/California/7/2009), shows a similar HI reactivity profile to the reference virus A/Bayern/69/2009, which carries G155E substitution in HA1. The effect of G155E substitution is less pronounced in the A/Lviv/N6/2009 reference virus, which contains an additional D222G HA1 substitution that is known to alter the receptor-binding properties of HA.

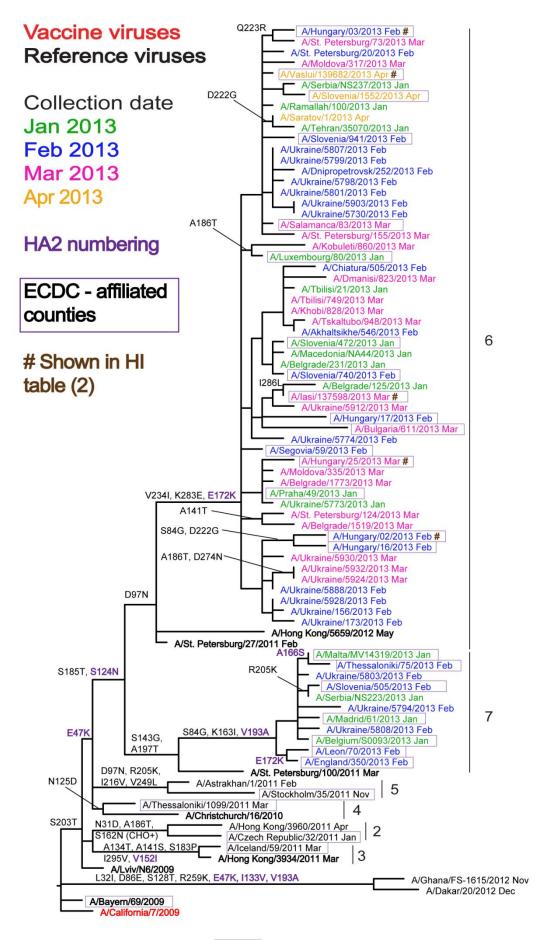
Phylogenetic analysis of the HA gene of representative viruses (Figure 1) shows that the H1N1 viruses from EU/EEA countries collected during the 2012–13 season cluster within genetic groups 6 and 7, with viruses belonging to group 6 predominating, notably so for those with collection dates since 1 March 2013. HA gene sequencing was performed on five test viruses, and their genetic grouping is shown in Table 2; all were in genetic aroup 6.

						Haemagglut	ination inhib	ition titre ¹				
			-			Post infe	ction ferret a	ntisera				1
/iruses		Collection	Passage	A/Cal	A/Bayern	A/Lviv	A/Chch	A/HK	A/St. P	A/St. P	A/HK	-
		date	History	7/09	69/09	N6/09	16/10	3934/11	27/11	100/11	5659/12	
				F30/11	F11/11	C4/34/09	F30/10	F21/11	F23/11	F24/11	F30/12	
	Genetic group						Group 4	Group 3	Group 6	Group 7	Group 6	
REFERENCE VIRUSES												1
VCalifornia/7/2009		2009-04-09	EP1/E2	320	320	640	160	160	160	320	160	
A/Bayern/69/2009		2009-07-01	MDCK5/MDCK1	80	160	80	40	<	40	40	40	G155E
A/Lviv/N6/2009		2009-10-27	MDCK4/S1/MDCK2	320	640	640	160	80	160	80	160	G155E, D2
VChristchurch/16/2010	4	2010-07-12	E2/E2	640	1280	1280	5120	640	1280	2560	640	
VHong Kong/3934/2011	3	2011-03-29	MDCK2/MDCK4	320	160	320	320	640	320	1280	640	
VSt. Petersburg/27/2011	6	2011-02-14	E1/E3	640	640	1280	640	640	1280	2560	1280	
VSt. Petersburg/100/2011	7	2011-03-14	E1/E2	640	640	640	640	1280	1280	2560	2560	
A/Hong Kong/5659/2012	6	2012-05-21	MDCK4/MDCK1	640	160	1280	640	1280	1280	2560	2560	
TEST VIRUSES												
A/Salaj/132819/2013		2013-01-24	MDCK2/C1	640	320	320	640	1280	1280	1280	1280	
VGalati/133254/2013		2013-01-31	MDCK1/C1	640	320	640	640	1280	1280	2560	1280	
VDolj/133772/2013	6	2013-02-04	MDCK1/C1	640	320	640	640	1280	1280	1280	1280	
VCluj/133922/2013	6	2013-02-06	MDCK1/C1	640	640	1280	640	1280	1280	2560	2560	
Vlasi/134271/2013		2013-02-12	MDCK1/C1	1280	320	640	640	1280	1280	5120	2560	
A/Hungary/02/2013	6	2013-02-12	MDCK2/E2/MDCK1	320	160	160	160	40	40	40	80	N156
VTimis/134705/2013	6	2013-02-15	MDCK2/C1	320	320	640	640	640	640	1280	1280	
A/Sibiu/134998/2013	6	2013-02-15	MDCK1/C1	640	640	1280	640	1280	1280	2560	2560	
Alasi/134898/2013		2013-02-18	MDCK1/C1	640	320	320	640	640	1280	2560	1280	
A/Hungary/03/2013	6	2013-02-20	MDCK1/E2/MDCK3	320	160	320	320	640	640	1280	1280	
VConstanta/135659/2013		2013-02-21	MDCK1/C1	640	320	640	640	1280	1280	2560	1280	
Alasi/135857/2013		2013-02-28	MDCK1/C1	640	320	640	320	640	640	1280	1280	
Alasi/137598/2013	6	2013-03-18	MDCK1/C1	1280	640	640	1280	1280	1280	5120	2560	
V/Hungary/25/2013	6	2013-03-18	MDCK2/E2/MDCK1	640	320	640	640	1280	1280	2560	2560	
	6	2013-04-02	MDCK1/C1	640	320	640	640	1280	1280	2560	1280	

Table 2. Antigenic analysis of A(H1N1)pdm09 viruses by HI

Sequences in phylogenetic tree (Figure 1)

Figure 1. Phylogenetic comparison of influenza A(H1N1)pdm09 HA genes



Influenza A(H3N2) virus analyses

Influenza A(H3N2) viruses continue to be difficult to characterise antigenically by HI assay due to variable agglutination of red blood cells from guinea pigs, turkeys and humans as described <u>before</u> [2]. The change in agglutination of red blood cells is associated with a reduced avidity of H3N2 viruses for the sialic acid receptor on the surface of the cell (<u>Lin et al. 2012</u>) [4]. Antigenic analyses of recently collected viruses conducted since the <u>May report</u> [1] are shown in Table 3. HI assays were carried out using guinea pig red blood cells in the presence of 20nM oseltamivir, added to circumvent the NA-mediated binding of H3N2 viruses to the red blood cells (<u>Lin et al. 2010</u>) [5]. The test viruses reacted poorly with post-infection ferret antiserum raised against the egg-propagated vaccine virus for 2012–13, A/Victoria/361/2011, compared with the titre against the homologous virus.

Generally, the test viruses also reacted poorly with antisera raised against other reference viruses and previous vaccine viruses propagated in eggs (A/Perth/16/2009, A/Victoria/208/2009, A/Iowa/19/2010 and A/Hawaii/22/2012). However, overall the panel of test viruses showed better reactivity with antiserum raised against egg-propagated A/Texas/50/2012 (the H3N2 vaccine virus <u>recommendation for the northern hemisphere</u> 2013–14) [6], compared with the titre of the antiserum with the homologous virus, than they did against other egg-propagated viruses. In Table 3, antiserum raised against A/Texas/50/2012 recognised 16 out of 19 test viruses at titres within fourfold of the titre to the homologous virus.

The test viruses reacted well with antisera raised against reference viruses exclusively propagated in in MDCK cells, and/or the derivative MDCK-SIAT-1 cells, when compared to the titres with the homologous viruses. These antisera were raised against cell-propagated virus isolates of A/Victoria/361/2011, A/Alabama/5/2010, A/Stockholm/18/2011, A/Berlin/93/2011 and A/Athens/112/2012.

Phylogenetic analysis of the HA gene sequences of representative viruses is shown in Figure 2. Viruses from EU/EEA countries collected since 1 January 2013 have HA genes that fall predominantly into genetic group 3C as is the case for all 13 test viruses sequenced during the preparation of this current report. Viruses carrying HA genes falling into group 3A and 3B (described in previous reports), 5 (e.g. A/Plzen/22/2013) and 6 (e.g. A/Lisboa/SU91/2012) have also been isolated earlier in the EU/EEA 2012–13 influenza season.

The amino acid substitutions in **HA1/HA2** associated with these groupings of recently collected viruses are:

- Group 3 viruses: N145S and V223I, with viruses in Groups 3B and 3C also carrying A198S, N312S; and in
- Group 3C: S45N (resulting in gain of a potential glycosylation site) and T48I, e.g. the prototype vaccine virus A/Victoria/361/2011; the great majority of viruses also carry the substitutions Q33R and N278K (e.g. A/Berlin/93/2011); an emerging subgroup also carries the substitutions T128A (resulting in the loss of a potential glycosylation site) and R142G;
- Group 3B: D158N;
- Group 3A: N144D, D158N;
- Group 5 viruses: D53N, Y94H, I230V and E280A (e.g. A/Alabama/05/2010), often in combination with K2E, N8D (resulting in the loss of a potential glycosylation site);
- Group 6 viruses: D53N, Y94H, S199A, I230V and E280A (e.g. A/Iowa/19/2010).

There is no evidence for antigenic change associated with any of the genetic groups or emerging subgroups, including the emerging subgroup in group 3C that carries substitutions in HA1 at amino acid residues 128 and 142.

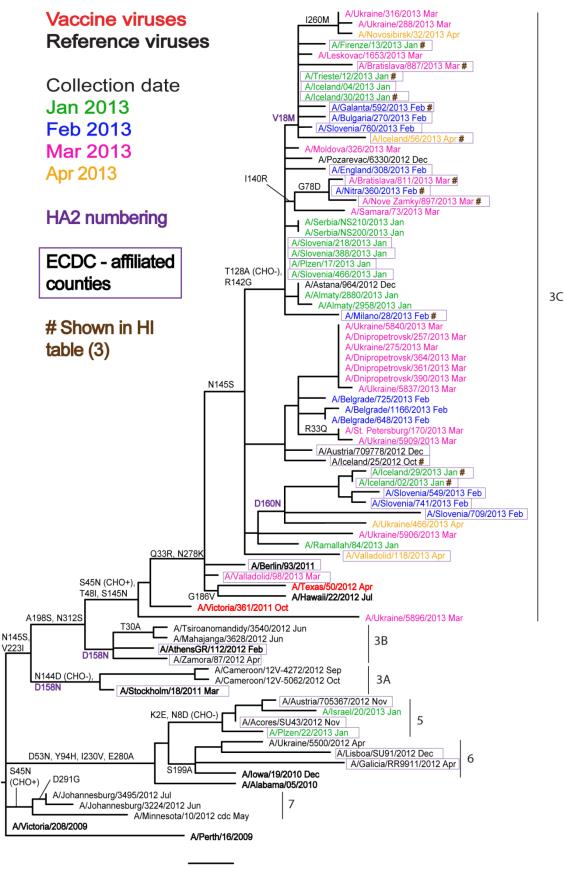
Table 3. Antigenic analysis of A(H3N2) viruses by HI (guinea pig RBC with 20nM oseltamivir)

								Haemaggl	utination inhit	oition titre ¹				
								Post inf	ection ferret a	ntisera				
Viruses		Collection Date	Passage History	A/Perth 16/09	A/Vic 208/09	A/Ala 5/10	A/Stock 18/11	A/lowa 19/10	A/Vic 361/11	A/Berlin 93/11	A/Vic 361/11	A/Athens 112/12	A/Texas 50/12	A/Hawa 22/1
				F35/11	F7/10	F27/10	F28/11	F15/11	Egg F35/12	T/C F11/12	T/C F14/12	F16/12	Egg F36/12	F37/*
	Genetic group					group 5	group 3A	group 6	group 3C	group 3C	group 3C	group 3B	group 3C	group 3
REFERENCE VIRUSES														
A/Perth/16/2009		2009-07-04	E3/E2	640	40	80	80	80	80	320	320	320	320	
A/Victoria/208/2009		2009-06-02	E3/E2	320	1280	320	640	1280	1280	2560	1280	1280	5120	25
A/Alabama/5/2010	5	2010-07-13	MK1/C2/SIAT2	<	<	80	80	80	80	160	320	160	160	
A/Stockholm/18/2011	3A	2011-03-28	SIAT5	40	80	80	160	80	160	320	320	320	640	3
A/lowa/19/2010	6	2010-12-30	E3/E2	320	640	640	1280	1280	640	2560	1280	1280	2560	12
A/Victoria/361/2011	3C	2011-10-24	E3/E2	320	640	320	80	640	2560	640	640	160	2560	e
A/Berlin/93/2011	3C	2011-12-07	NVD3/SIAT6	80	80	320	320	160	160	640	640	640	1280	3
A/Victoria/361/2011	3C	2011-10-24	MDCK2/SIAT2	40	80	80	160	80	160	640	640	640	320	1
A/Athens/112/2012	3B	2012-02-01	SIAT7	80	80	160	320	160	320	640	640	1280	1280	
A/Texas/50/2012	3C	2012-04-15	E5/E1	320	640	320	1280	640	1280	2560	1280	1280	2560	1:
A/Hawaii/22/2012	3C	2012-07-09	E4/E1	320	640	320	640	1280	640	2560	1280	1280	2560	51
TEST VIRUSES														
A/Iceland/25/2012	3C	2012-10-17	MDCK1/SIAT3	40	80	160	320	160	160	640	640	640	640	:
A/Iceland/27/2012		2012-10-17	MDCK2/SIAT1	40	80	80	160	80	160	640	320	320	320	
A/Iceland/02/2013	3C	2013-01-02	MDCKx/SIAT1	40	40	80	160	80	160	640	640	640	640	:
A/Iceland/14/2013		2013-01-10	MDCK1/SIAT1	40	80	80	320	80	160	640	640	640	640	:
A/lceland/29/2013	3C	2013-01-24	MDCK1/SIAT3	40	40	40	160	40	160	640	320	640	640	:
A/Firenze/13/2013	3C		MDCK2/SIAT1	<	80	80	160	80	160	640	640	640	640	:
A/Parma/27/2013		2013-01-25	MDCK2/SIAT2	<	80	80	160	80	160	640	320	640	640	:
A/Perugia/14/2013		2013-01-27	MDCK2/SIAT2	<	40	80	160	80	160	640	320	640	640	:
A/Perugia/13/2013		2013-01-28	MDCK1/SIAT1	<	80	80	160	80	160	640	640	640	640	3
A/Trieste/12/2013	3C	2013-01-28	SIAT2	<	80	80	160	80	160	1280	320	640	640	3
A/Iceland/30/2013	3C	2013-01-28	MDCKx/SIAT1	<	40	40	160	40	80	320	320	320	320	-
A/Nitra/360/2013	3C	2013-02-05	MDCK1/SIAT1	80	160	160	320	160	320	1280	640	1280	1280	
A/Bratislava/530/2013	_	2013-02-15	MDCK2/SIAT1	80	160	160	320	320	320	1280	1280	640	1280	e
A/Milano/28/2013	3C	2013-02-19	MDCK1/SIAT3	<	40	40	160	80	80	640	320	320	320	1
A/Galanta/592/2013	3C	2013-02-21	MDCK1/SIAT1	40	80	80	160	80	160	640	320	640	640	
A/Bratislava/811/2013	3C	2013-03-07	MDCK1/SIAT1	80	320	320	640	320	640	1280	1280	1280	2560	e
A/Bratislava/887/2013	3C	2013-03-14	MDCK1/SIAT1	80	160	320	320	160	160	1280	1280	1280	1280	
A/Nove Zamky/897/2013	3C	2013-03-15	MDCK1/SIAT1	320	640	640	1280	640	1280	2560	2560	2560	2560	2
A/Iceland/56/2013	3C	2013-04-26	MDCK1/SIAT1	80	80	160	320	80	160	640	640	320	640	3

Sequences in phylogenetic tree (Figure 2)

Vaccine 2012-2013 Vaccine 2013-2014

Figure 2. Phylogenetic comparison of influenza A(H3N2) HA genes





Influenza B virus analyses

B/Victoria-lineage viruses

Table 4 shows the results of antigenic analyses for viruses of the B/Victoria-lineage performed since the May report [1]. All test viruses were from Hungary. Compared with the titre against the homologous virus in HI assays only one test virus, B/Hungary/05/2013, showed poor reactivity with post-infection ferret antiserum raised against the eqq-propagated virus B/Brisbane/60/2008, a component of trivalent vaccines for the 2010-11 season and a recommended component of quadrivalent vaccines [6] for the 2013–14 northern hemisphere influenza season.

B/Hungary/05/2013 showed similarly reduced reactivities with antisera raised against other reference viruses propagated in hens' eggs: B/England/393/2008, B/Malta/636714/2011 and B/Johannesburg/3964/2012. These observations probably relate to the loss of an N-linked carbohydrate site at position 197 of HA1 which is commonly associated with growth of B/Victoria-lineage viruses in hens' eggs, resulting in the exposure of a dominant antigenic site. Sequencing for four of the Hungarian viruses is complete: 05/2013 has retained the glycosylation site (NET), while 07/2013, 23/2013 and 26/2013 have lost it due to amino acid substitution or polymorphism (NEA, NEI and XEX respectively). All test viruses showed a consistent reactivity pattern, with titres close to the homologous titres, for antisera raised against reference viruses genetically closely related to B/Brisbane/60/2008 but propagated in cells; these post-infection ferret antisera were raised against B/Paris/1762/2008, B/Hong Kong/514/2009, B/Odessa/3886/2010 and B/Formosa/V2367/2012.

Phylogenetic analysis of the HA genes of representative B/Victoria-lineage viruses is shown in Figure 3. All the viruses received with collection dates in 2013 from EU/EEA laboratories carried HA genes that fell into genetic clade 1A. The amino acid substitution associated with the separation of clade 1 into clades 1A and 1B, L58P, has no apparent effect on antigenicity. The HAs of recent viruses show few amino acid substitutions compared with B/Brisbane/60/2008.

						Haemag	glutination	inhibition tit	re				
							Post infe	ction ferret a	ntisera				
Viruses		Collection date	Passage History	B/Bris ² 60/08	B/Mal ¹ 2506/05	B/Eng ¹ 393/08	B/Bris ¹ 60/08	B/Paris ¹ 1762/08	B/HK ¹ E 514/09	3/Odessa ¹ 3886/10	B/Malta ¹ 636714/11	B/Jhb ¹ 3964/12	B/Foi V2367/1
				Sh 522	F37/11	F05/11	F22/12	F07/11	F13/10	F19/11	F33/11	F01/13	F04/1
	Genetic group					1A	1A	1A	1B	1B	1A	1A	1A
REFERENCE VIRUSES													
B/Malaysia/2506/2004		2004-12-06	E3/E6	1280	320	40	80	<	<	<	80	160	
B/England/393/2008	1A	2008-08-29	E1/E2	2560	40	320	320	80	40	40	160	320	1
B/Brisbane/60/2008	1A	2008-08-04	E4/E3	1280	80	320	320	80	40	40	160	320	1
B/Paris/1762/2008	1A	2009-02-09	C2/MDCK2	2560	<	20	40	80	80	40	20	40	
B/Hong Kong/514/2009	1B	2009-10-11	MDCK4	2560	<	10	20	80	80	80	10	40	
B/Odessa/3886/2010	1B	2010-03-19	C2/MDCK2	2560	40	160	160	80	40	40	80	320	3
B/Malta/636714/2011	1A	2011-03-07	E4/E1	2560	80	320	320	80	40	40	320	320	3
B/Johannesburg/3964/2012	1A	2012-08-03	E1/E1	2560	160	640	640	80	80	40	320	1280	6
B/Formosa/V2367/2012	1A	2012-08-06	MDCK1/MDCK2	1280	10	80	80	80	40	40	80	160	1
TEST VIRUSES													
B/Hungary/06/2013		2013-02-25	MDCK2/E2/MDCK2	2560	160	160	320	80	40	40	160	320	3
B/Hungary/07/2013	1A	2013-02-27	MDCK2/E3/MDCK3	2560	40	320	320	80	40	40	160	320	3
B/Hungary/05/2013	1A	2013-02-27	MDCK2/E2/MDCK1	5120	40	40	40	80	80	80	20	40	
B/Hungary/13/2013		2013-03-13	MDCK2/E2/MDCK1	2560	80	160	160	40	20	20	160	320	1
B/Hungary/23/2013	1A	2013-04-02	E2/E1	1280	160	80	160	40	20	40	80	160	1
B/Hungary/26/2013	1A	2013-04-08	MDCK1/E1/MDCK2	1280	80	160	160	40	20	20	160	160	3

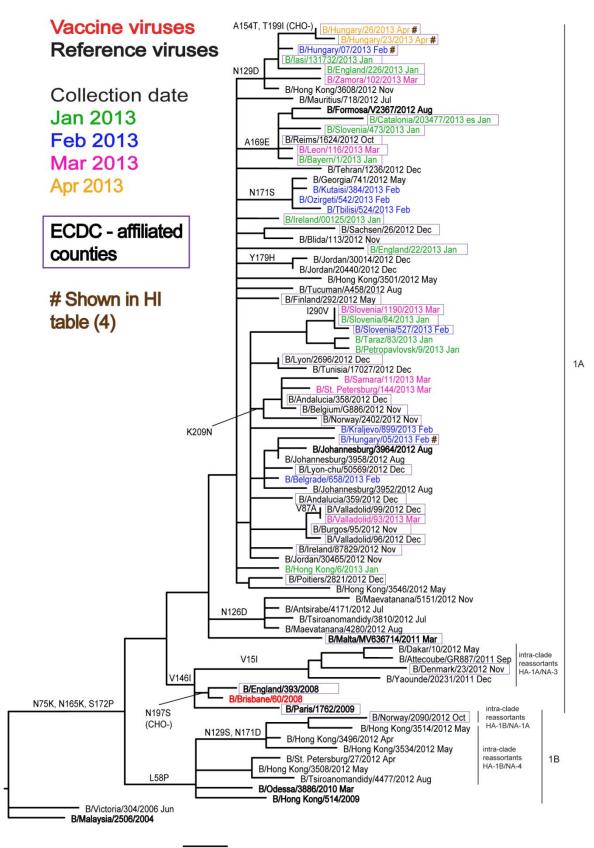
Table 4. Antigenic analysis of influenza B viruses (Victoria-lineage) by HI

1. < = <10; 2. hyperimmune sheep serum

Sequences in phylogenetic analysis (Figure 3)

Recommended B-Victoria lineage component for quadravalent vaccine 2012-13 and 2013-14

Figure 3. Phylogenetic comparison of influenza B/Victoria-lineage HA genes



9.0E-4

B/Yamagata-lineage viruses

Table 5 shows the results of HI analyses of B/Yamagata lineage viruses tested since the May report [1]. The genetic clade into which sequenced HA genes of test viruses fall is indicated.

Eleven of 12 test viruses showed good reactivity (within fourfold of the homologous titre) with antisera raised against the egg-propagated vaccine virus recommended for the northern hemisphere winter 2013–14 influenza season [6], B/Massachusetts/02/2012. Antiserum raised against egg-propagated B/Wisconsin/1/2010, the virus used in the vaccine for 2012–13, also showed reactivity within fourfold of the titre against the homologous virus for nine of the test viruses. B/Hungary/22/2013 showed low reactivity with all antisera in the test panel, and in terms of HA sequence its only unusual feature was the loss of a potential N-linked carbohydrate site at position 196 of HA1 (NKT \rightarrow NKN), compared to the HA sequences of other test viruses. Antisera raised against cell-propagated viruses, whether of clade 2 or clade 3, showed good reactivity (within fourfold of the homologous titre) against the majority of test viruses. Seven of the test viruses had been genetically characterised at the time of preparation of this report, with all falling into genetic group 2.

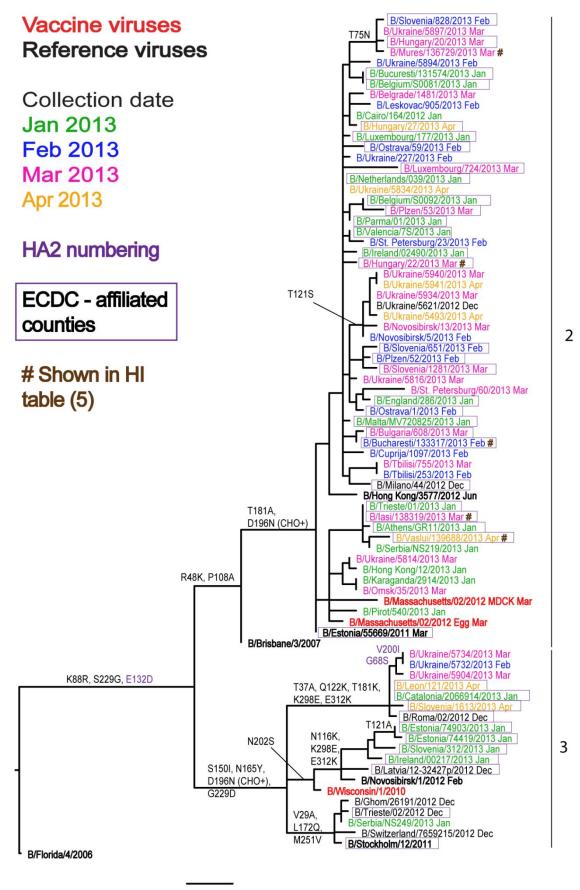
Figure 4 shows a phylogenetic analysis of the HA genes of representative B/Yamagata-lineage viruses. The analysis shows that the HA genes of recent viruses continue to fall into two genetic clades: clade 3 (represented by the vaccine virus B/Wisconsin/1/2010 and reference viruses B/Stockholm/12/2011 and B/Novosibirsk/1/2012) and clade 2 (represented by the reference viruses B/Brisbane/3/2007, B/Estonia/55669/2011, B/Hong Kong/3577/2012 and the 2013–14 vaccine virus B/Massachusetts/02/2012). The two clades are differentiated by substitutions at HA1 residues 48, 108, 150, 165, 181 and 229. The HA genes of viruses of clade 2 encode K48, A108, S150. N165, A181 and G229: the HA genes of viruses in clade 3 encode R48, P108, I150, Y165, T181 and D229. The proportion of viruses received with HA genes that fall into clade 2 has continued to increase over the number with HA genes falling into clade 3.

				-				Po	at infection ferre	t antisera			
Viruses	Genetic group	Collection date	Passage History	B/FI ³ 4/06 SH479	B/FI ¹ 4/06 F1/10 Group 1	B/Bris ² 3/07 F21/12 Group 2	B/Wis ² 1/10 F24/12 Group 3	B/Stock ² 12/11 F12/12 Group 3	B/Estonia ² 55669/11 F26/11 Group 2	B/Novo ² 1/12 F31/12 Group 3	B/HK ² 3577/12 F33/12 Group 2	B/Mass ² 2/12 Egg F07/13 Group 2	B/Mass 2/1 T/C F08/1 Group
REFERENCE VIRUSES													
B/Florida/4/2006	1	2006-12-15	E7/E1	5120	1280	1280	320	640	320	80	640	1280	8
B/Brisbane/3/2007	2	2007-09-03	E2/E2	2560	640	640	320	320	160	80	320	640	8
B/Wisconsin/1/2010	3	2007-08-07	E3/E2	1280	640	320	320	320	20	80	80	320	4
B/Stockholm/12/2011	3	2007-08-07	E4/E1	1280	160	160	80	160	<	40	40	160	2
B/Estonia/55669/2011	2	2011-03-14	MDCK2/MDCK3	1280	160	80	160	160	640	160	640	160	32
B/Novosibirsk/1/2012	3	2012-02-14	C2/MDCK3	2560	320	160	640	320	320	320	320	160	64
B/Hong Kong/3577/2012	2	2012-06-13	MDCK4	2560	160	160	320	160	640	160	640	160	32
B/Massachusetts/02/2012	2	2012-03-13	E3/E3	2560	640	640	640	320	80	40	160	640	4
B/Massachusetts/02/2012	2	2012-03-13	MDCK1/C2/MDCK2	1280	160	160	80	80	160	40	320	160	8
TEST VIRUSES													
B/Bucuresti/133317/2013	2	2013-02-01	MDCK1/MDCK1	1280	160	20	40	80	640	40	320	160	16
B/Sibiu/133676/2013		2013-02-04	MDCK1/MDCK1	5120	320	80	160	320	640	160	640	320	64
B/Suceava/133820/2013	2	2013-02-06	MDCK1/MDCK1	1280	160	40	80	80	640	80	320	160	32
B/Hungary/04/2013		2013-02-15	MDCK1/E3/MDCK2	1280	160	160	80	160	320	40	320	160	8
B/lasi/135846/2013		2013-02-27	MDCK1/MDCK1	1280	320	40	80	80	320	80	640	160	32
B/Bihor/135892/2013	2	2013-02-27	MDCK1/MDCK1	1280	320	40	80	10	640	40	320	160	32
B/Hungary/15/2013		2013-03-04	MDCK2/E2/MDCK2	2560	160	160	80	160	1280	80	640	160	32
B/Mures/136729/2013	2	2013-03-06	MDCK1/MDCK1	1280	160	40	40	160	640	80	320	160	32
B/Hungary/22/2013	2	2013-03-26	E3/E1	320	80	40	10	20	40	<	40	80	2
B/lasi/138319/2013	2	2013-03-26	MDCK1/MDCK1	2560	320	80	160	160	640	160	640	320	64
		2013-03-26	MDCK1/MDCK1	1280	160	160	80	160	640	80	640	160	16
B/lasi/138316/2013				0500	640	80	160	320	640	160	640	320	64
B/lasi/138316/2013 B/Vaslui/139688/2013	2	2013-04-08	MDCK1/MDCK1	2560	640	80	100	520	040	100	040	520	04

Table 5. Antigenic analysis of influenza B viruses (Yamagata lineage) by HI

Sequences in phylogenetic analysis (Figure 4)

Figure 4. Phylogenetic comparison of influenza B/Yamagata-lineage HA genes



0.002

Influenza A(H7N9) virus

On 1 April 2013, the <u>WHO Global Alert and Response</u> [7] reported that the China Health and Family Planning Commission notified the World Health Organization (WHO) of three cases of human infection with influenza A(H7N9). The cases were confirmed by laboratory testing on 29 March by the Chinese CDC. A description of the characteristics of H7N9 viruses can be found on the <u>WHO website</u> [8]. WHO is updating information on the outbreak <u>regularly</u> [8] and ECDC is posting <u>epidemiological updates</u> [9]. A <u>Rapid Risk Assessment</u> [10] for these A(H7N9) viruses has been carried out and posted by ECDC on 3 April 2013, and an updated risk assessment has been posted by <u>WHO</u> [11]. As of 4 July 2013, <u>WHO</u> reported [12] 133 laboratory confirmed cases and 43 associated fatalities.

A description of results generated by the WHO Collaborating Centre for Reference and Research on Influenza at the MRC National Institute for Medical Research in London, and evaluated at the WHO Vaccine Composition Meetings held in Beijing, China, on 17–19 September 2012 and at WHO Geneva on 18–20 February 2013, can be found at:

http://www.nimr.mrc.ac.uk/documents/about/Interim_Report_September_2012_2.pdf [14]

http://www.nimr.mrc.ac.uk/documents/about/Interim_Report_February 2013.pdf [15]

Note on the figures

The phylogenetic trees were constructed using RAxML and drawn using *FigTree*. The bars indicate the proportion of nucleotide changes between sequences. Reference strains are viruses to which post-infection ferret antisera have been raised. The colours indicate the month of sample collection. Isolates from WHO NICs in ECDC countries are highlighted within boxes. Sequences for some of the viruses from non-EU/EEA countries were recovered from GISAID. We acknowledge all laboratories who submitted sequences directly to the London WHO Collaborating Centre.

References

1 European Centre for Disease Prevention and Control. Influenza virus characterisation – Summary Europe, May 2013. Stockholm: ECDC; 2013 [cited 2013 Jun 1]. Available from:

http://www.ecdc.europa.eu/en/publications/Publications/influenza-virus-characterisation-May-2013.pdf

2 European Centre for Disease Prevention and Control. Influenza virus characterisation – Summary Europe, April 2013. Stockholm: ECDC; 2013 [cited 2013 Jun 1]. Available from: http://www.ecdc.europa.eu/en/publications/Publications/influenza-virus-characterisation-April-2013.pdf

3 European Centre for Disease Prevention and Control. Influenza virus characterisation - Summary Europe, April 2011. Stockholm: ECDC; 2011 [cited 2013 Jun 1]. Available from: http://ecdc.europa.eu/en/publications/Publications/1304 Influenza virus characterisation 2011 April.pdf

4 Lin YP, Xiong X, Wharton SA, Martin SR, Coombs PJ, Vachieri SG, et al. Evolution of the receptor binding properties of the influenza A(H3N2) hemagglutinin. Proc Natl Acad Sci USA. 2012 Dec 26;109(52):21474-9.

5 Lin YP, Gregory V, Collins P, Kloess J, Wharton S, Cattle N, et al. Neuraminidase receptor binding variants of human influenza A(H3N2) viruses resulting from substitution of aspartic acid 151 in the catalytic site: a role in virus attachment? J Virol. 2010 Jul;84(13):6769-81. Available from:

http://jvi.asm.org/cgi/content/full/84/13/6769?view=long&pmid=20410266

6 World Health Organization, Weekly epidemiological record [serial on the internet], 2013 [cited 2013 Jun 1];88(10) Available from: http://www.who.int/wer/2013/wer8810.pdf

7 World Health Organization. Human infection with influenza A(H7N9) virus in China. Global Alert and Response (GAR) [serial on the internet]. Apr 1 2013 [cited 2013 June 1]. Available from: http://www.who.int/csr/don/2013 04 01/en/index.html

8 World Health Organization. Avian influenza A(H7N9) virus. [homepage on the Internet]. 2013 [cited 2013 June 20]. Available from: http://www.who.int/influenza/human animal interface/influenza h7n9/en/index.html

9 European Centre for Disease Prevention and Control. Epidemiological updates. [homepage on the Internet]. 2013 [cited 2013 Jun 12]. Available from:

http://ecdc.europa.eu/en/press/epidemiological updates/Pages/epidemiological updates.aspx

10 European Centre for Disease Prevention and Control. Severe respiratory disease associated with a novel influenza A virus, A(H7N9) – China, 3 April 2013. Stockholm: ECDC; 2013 [cited 2013 Jun 12]. Available from: http://ecdc.europa.eu/en/publications/Publications/AH7N9-China-rapid-risk-assessment.pdf

11 World Health Organization. WHO risk assessment: Human infections with avian influenza A(H7N9) virus, 7 June 2013. Geneva: WHO; 2013 [cited 2013 Jun 12]. Available from:

http://www.who.int/influenza/human_animal_interface/influenza_h7n9/RiskAssessment_H7N9_07Jun13.pdf

13 World Health Organization. Human infection with influenza A(H7N9) virus in China. Global Alert and Response (GAR) [serial on the internet]. Apr 1 2013 [cited 2013 June 1]. Available from: http://www.who.int/csr/don/2013 07 04/en/index.html

14 National Institute for Medical Research, WHO Influenza Centre London. Report prepared for the WHO annual consultation on the composition of influenza vaccine for the southern hemisphere 2013, 17th-19th September 2012. London: WHO Collaborating Centre for Reference and Research on Influenza, National Institute for Medical Research: 2012. Available from:

http://www.nimr.mrc.ac.uk/documents/about/Interim_Report_September_2012_2.pdf

15 National Institute for Medical Research, WHO Influenza Centre London, Report prepared for the WHO annual consultation on the composition of influenza vaccine for the northern hemisphere 2013/14, 18th–20th February 2013. London: WHO Collaborating Centre for Reference and Research on Influenza, National Institute for Medical Research; 2013. Available from: http://www.nimr.mrc.ac.uk/documents/about/Interim Report February 2013.pdf