# Influenza virus characterisation 

Summary Europe, May 2016

## Summary

From week 40/2015, the start of weekly reporting on influenza activity in the WHO European Region, to week 20/2016 over 138000 influenza detections across the Region have been reported. Influenza type A viruses have prevailed over type B but, unlike the situation in the 2014-15 season, $A(H 1 N 1)$ pdm09 viruses greatly outnumbered $A(H 3 N 2)$ and the proportion of $B /$ Victoria-lineage detections has risen substantially, representing $\sim 92 \%$ of those assigned to a lineage.
To date, 26 EU/EEA countries have shared 631 influenza-positive specimens with the Francis Crick Institute, London, for detailed characterisation: one additional country and 31 specimens since the April 2016 report. Since the latter report, 75 viruses have been characterised antigenically, and genetic analyses are ongoing.
All of $49 \mathrm{~A}(\mathrm{H} 1 \mathrm{~N} 1)$ pdm09 viruses characterised antigenically were similar to the vaccine virus $A / C a l i f o r n i a / 7 / 2009$. Worldwide new genetic subclusters of viruses within the 6B clade have emerged, with two being designated as subclades: 6B.1 defined by HA1 amino acid substitutions S162N and I216T and 6B. 2 defined by HA1 amino acid substitutions V152T and V173I. Of the 246 viruses characterised genetically for the 2015-16 season, 25 (10\%) were clade 6B, 214 (87\%) were subclade 6B. 1 and seven (3\%) were subclade 6B.2.
The three $A(H 3 N 2)$ test viruses characterised by haemagglutination inhibition (HI) assay were poorly recognised by reference antiserum raised against egg-propagated A/Switzerland/9715293/2013, the vaccine virus recommended for use in the 2015-2016 northern hemisphere influenza season. The test viruses were recognised somewhat better by antisera raised against egg-propagated $A / H o n g$ Kong/4801/2014, the virus recommended for use in 2016 southern hemisphere and 2016-2017 northern hemisphere influenza vaccines. Of 65 A(H3N2) viruses characterised genetically for the 2015-2016 season, one (1\%) was clade 3C.3, 40 ( $62 \%$ ) were subclade 3C. 2 a and 24 ( $37 \%$ ) were subclade 3C.3a.

The $22 \mathrm{~B} /$ Victoria-lineage viruses were antigenically similar to tissue culture-propagated surrogates of B/Brisbane/60/2008. All 78 viruses characterised genetically for the 2015-2016 season fell in genetic clade 1 A , as do recently collected viruses worldwide.

One $B /$ Yamagata virus have been characterised since the previous report; it reacted well with post-infection ferret antiserum raised against egg-propagated $B /$ Phuket/3073/2013, the recommended vaccine virus for the northern hemisphere 2015-16 influenza season and for quadrivalent vaccines in the 2016 southern hemisphere and 2016-17 northern hemisphere seasons. All 16 viruses characterised genetically for the 2015-2016 season fell in genetic clade 3.

[^0]Table 1 shows a summary of influenza virus detections in the WHO European Region reported to The European Surveillance System (TESSy) for the first 34 weeks (weeks 40/2015-20/2016) of reporting for the 2015-16 season. A total of over 138000 detections had been made with type A viruses prevailing over type $B$ at a ratio of 2.3:1; this compares to a ratio of $5.8: 1$ to week $7 / 2016$ indicating a surge in influenza type $B$ circulation over the last 13 weeks. So far, of the type A viruses subtyped ( $n=66707$ ) and the type $B$ viruses ascribed to lineage ( $n=$ 7 834), $A(H 1 N 1)$ pdm09 have prevailed over $A(H 3 N 2)$ and $B / V i c t o r i a ~ o v e r ~ B / Y a m a g a t a ~ b y ~ r a t i o s ~ o f ~ 10.2: 1 ~ a n d ~$ 11.1:1, respectively.

Since the start of weekly reporting for the 2015-16 influenza season (week 40/2015), 47 shipments of specimens have been received at the Crick Worldwide Influenza Centre (WIC), from 26 countries in the EU/EEA (Table 2). Of the 631 specimens received - a mix of clinical samples and virus isolates - the majority ( $79 \%$ ) were type A viruses, and $A(H 1 N 1)$ pdm09 outnumbered $A(H 3 N 2)$ at a ratio of 5.2:1. Of the 134 type $B$ specimens received ( $21 \%$ of the specimens), 114 were B/Victoria-lineage and 16 B/Yamagata-lineage. A number of specimens are still in the process of being characterised. The antigenic and genetic properties of influenza virus isolates characterised since the April 2016 report $^{1}$ are presented and discussed in this report.
Table 1. Influenza virus detections in the WHO European Region since the start of reporting for the 2015-16 season (weeks 40/2015-20/2016) ${ }^{\text {a }}$

| Virus type/subtype | Cumulative number of detections |  |  | Totals* |  |
| :---: | :---: | :---: | :---: | :---: | :---: |
|  | Sentinel sources | Non-sentinel sources | Totals | \% | Ratios |
| Influenza A | 10496 | 85919 | 96415 | 69.7 | 2.3:1 |
| A(H1N1)pdm09 | 8665 | 52083 | 60748 | 91.1 | 10.2:1 |
| A(H3N2) | 1365 | 4594 | 5959 | 8.9 |  |
| A not subtyped | 466 | 29242 | 29708 |  |  |
| Influenza B | 8144 | 33791 | 41935 | 30.3 |  |
| Victoria lineage | 3974 | 3210 | 7184 | 91.7 | 11.1:1 |
| Yamagata lineage | 145 | 505 | 650 | 8.3 |  |
| Lineage not ascribed | 4025 | 30076 | 34101 |  |  |
| Total detections (total tested) | 18640 (50 861) | 119710 (536 625) | 138350 (587 486) |  |  |

* Percentages are shown for total detections (types A \& B, and for viruses ascribed to subtype/lineage). Ratios are given for type $A: B, A(H 1 N 1) p d m 09: A(H 3 N 2)$ and Victoria:Yamagata lineages.

[^1][^2]Table 2. Summary of clinical samples and virus isolates received from EU/EEA Member States: packages received since the start of the 2015-16 reporting period (week 40/2015)


* Month indicates the months in which the clinical specimens were collected

1. Propagated to sufficient titre to perform HI assay
2. Propagated to sufficient titre to perform HI assay in presence of 20 nM oseltamivir; numbers in red indicate viruses recovered but with insufficient HA titre to permit HI assay

## Influenza A(H1N1)pdm09 virus analyses

Haemagglutination inhibition (HI) analyses of viruses that have been performed since the April 2016 report are shown in Tables 3-1 to 3-3. All of the 49 A(H1N1)pdm09 viruses from EU/EEA countries characterised antigenically were similar to the vaccine virus, A/California/7/2009. Generally, the test viruses were recognised by the panel of antisera at titres within fourfold of the titres for the homologous viruses, with the exception of the antiserum raised against $A / C h r i s t c h u r c h / 16 / 2010$. This antiserum recognised only $3 / 49(6 \%)$ test viruses at a titre within fourfold of the titre for the homologous virus. In addition, antiserum raised against A/Lviv/N6/2009 showed eightfold reduced titres with five of the test viruses, compared with the homologous titres. Reference viruses carrying HA1 G155E amino acid substitutions, A/Bayern/69/2009 and A/Lviv/N6/2009, showed reduced recognition by the antisera raised against A/California/7/2009 and reference viruses in genetic clades 4, 5, 6, 7 and subclades $6 A, 6 B, 6 B .1$ and 6B.2.
While sequencing is ongoing for many of the test viruses indicated in Tables 3-1 to 3-3, the 27 viruses characterised genetically all fell in subclade 6B. 1 (Tables 3-1 \& 3-2). Since 2009, the HA genes have evolved, and nine clades have been designated. For well over a year, viruses in clade 6, represented by A/St Petersburg/27/2011 and carrying amino acid substitutions of D97N, S185T and S203T in HA1 and E47K and S124N in HA2 compared with A/California/7/2009, have predominated worldwide with a number of subclades emerging. All EU/EEA viruses characterised since the September 2014 report ${ }^{2}$ carry HA genes in subclade 6B, which is characterised by additional amino acid substitutions of K163Q, A256T and K283E in HA1 and E172K in HA2 compared with A/California/7/2009, e.g. A/South Africa/3626/2013. A number of virus clusters have emerged within clade 6B, and two of these have been designated as subclades: viruses in subclade 6B. 1 are defined by HA1 amino acid substitutions S84N, S162N (which results in the formation of a new potential glycosylation motif at residues 162-164 of HA1) and I216T, while those in subclade 6B. 2 are defined by HA1 amino acid substitutions V152T and V173I (Figure 1).

[^3]Table 3-1. Antigenic analysis of A(H1N1)pdm09 viruses by HI


| Viruses | Other <br> information |  | Collectiondate | Passage history | Haemagglutination inhibition titre |  |  |  |  |  |  |  |  |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
|  |  |  |  |  | Post-infection ferret antisera |  |  |  |  |  |  |  |  |  |  |
|  |  |  |  |  | A/Cal | A/Bayern | A/Lviv | A/Chch | A/Astrak | ASSt. P | A/St. P | A/HK | A/Sth Afr | A/Slov | Alsrael |
|  |  |  |  |  | 7/09 | 69/09 | N6/09 | 16/10 | 1/11 | 27/11 | 100/11 | 5659/12 | 3626/13 | 2903/2015 | Q-504/15 |
|  |  | Passage history |  |  | Egg | MDCK | MDCK | Egg | mDCK | Egg | Egg | MDCK | Egg | Egg | mDCK |
|  |  | number |  |  | F06/16 ${ }^{\text {¹ }}$ | F09/15 ${ }^{\text {¹ }}$ | F14/13 ${ }^{\text {+ }}$ | F15/4 ${ }^{4}$ | F22/13 ${ }^{\text {4 }}$ | F26/14 ${ }^{\text {¹ }}$ | F24/11 ${ }^{11}$ | F30/12 ${ }^{\text {-1 }}$ | F03/14 ${ }^{\text {a }}$ | F02/16 ${ }^{\text {2 }}$ | F08/16 ${ }^{\text {2 }}$ |
|  |  | Genetic group |  |  |  |  |  | 4 | 5 | 6 | 7 | 6 A | 6 B | 6B. 1 | 6B. 2 |
| REFERENCE VIRUSES |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |
| A/California/7/2009 | clone 38-32 |  | 2009-04-09 | E3/E2 | 2560 | 1280 | 640 | 640 | 2560 | 640 | 2560 | 2560 | 1280 | 2560 | 2560 |
| A/Bayern/69/2009 |  |  | 2009-07-01 | MDCK5/MDCK1 | 80 | 320 | 320 | 80 | 40 | 40 | 80 | 40 | 80 | 40 | 40 |
| A/Lviv/N6/2009 |  |  | 2009-10-27 | MDCK4/SIAT $1 / \mathrm{MDCK} 3$ | 160 | 1280 | 640 | 320 | 160 | 80 | 160 | 320 | 160 | 320 | 160 |
| A/Christchurch/16/2010 |  | 4 | 2010-07-12 | E1/E3 | 1280 | 1280 | 1280 | 5120 | 2560 | 640 | 2560 | 1280 | 1280 | 2560 | 1280 |
| A/Astrakhan/1/2011 |  | 5 | 2011-02-28 | MDCK1/MDCK5 | 1280 | 640 | 320 | 640 | 1280 | 640 | 2560 | 1280 | 1280 | 1280 | 1280 |
| A/St. Petersburg/27/2011 |  | 6 | 2011-02-14 | E1/E4 | 1280 | 640 | 320 | 640 | 1280 | 640 | 2560 | 1280 | 1280 | 2560 | 1280 |
| A/St. Petersburg/100/2011 |  | 7 | 2011-03-14 | E1/E4 | 1280 | 1280 | 640 | 640 | 1280 | 640 | 2560 | 1280 | 640 | 1280 | 1280 |
| A/Hong Kong/5659/2012 |  | 6 A | 2012-05-21 | MDCK4/MDCK2 | 640 | 320 | 160 | 160 | 320 | 160 | 1280 | 640 | 320 | 640 | 320 |
| A/South Africa/3626/2013 |  | 6 B | 2013-06-06 | E1/E3 | 640 | 320 | 320 | 640 | 640 | 320 | 1280 | 640 | 640 | 640 | 640 |
| A/SIovenia/2903/2015 | clone 37 | 6 B .1 | 2015-10-26 | E4/E1 | 2560 | 640 | 640 | 1280 | 1280 | 640 | 5120 | 2560 | 1280 | 2560 | 1280 |
| A/lsrael/Q-504/2015 |  | 6B. 2 | 2015-12-15 | C1/MDCK2 | 1280 | 640 | 320 | 640 | 1280 | 640 | 2560 | 1280 | 1280 | 2560 | 2560 |
| test viruses |  |  |  |  |  |  |  |  |  |  |  |  |  |  |  |
| A/Bulgaria/404/2016 |  | 68. 1 | 2016-02-09 | SIAT2/MDCK1 | 1280 | 640 | 320 | 640 | 1280 | 640 | 2560 | 1280 | 1280 | 2560 | 2560 |
| A/Bulgaria/359/2016 |  | 6B. 1 | 2016-02-09 | SIAT1/MDCK1 | 1280 | 640 | 320 | 640 | 640 | 640 | 1280 | 1280 | 1280 | 2560 | 1280 |
| A/Bulgaria/333/2016 |  | 68. 1 | 2016-02-09 | SIAT2/MDCK1 | 1280 | 640 | 320 | 640 | 640 | 640 | 1280 | 1280 | 1280 | 2560 | 2560 |
| A/Bulgaria/332/2016 |  | 6 B .1 | 2016-02-09 | SIAT2/MDCK1 | 1280 | 640 | 640 | 1280 | 1280 | 640 | 2560 | 2560 | 1280 | 2560 | 2560 |
| A/Bulgaria/381/2016 |  | 6 B .1 | 2016-02-10 | SIAT1/MDCK1 | 1280 | 640 | 320 | 640 | 640 | 320 | 1280 | 1280 | 1280 | 1280 | 1280 |
| A/Bulgaria/507/2016 |  | 6B. 1 | 2016-02-13 | SIAT1/MDCK1 | 1280 | 640 | 320 | 640 | 1280 | 320 | 1280 | 1280 | 1280 | 2560 | 2560 |
| A/Bulgaria/505/2016 |  | 6 B .1 | 2016-02-13 | SIAT1/MDCK1 | 1280 | 640 | 320 | 640 | 1280 | 640 | 1280 | 1280 | 1280 | 2560 | 2560 |
| A/Bulgaria/531/2016 |  | 6 B .1 | 2016-02-16 | SIAT1/MDCK1 | 1280 | 640 | 320 | 640 | 1280 | 640 | 2560 | 1280 | 1280 | 1280 | 1280 |
| A/Bulgaria/559/2016 |  | 6B. 1 | 2016-02-17 | SIAT2/MDCK1 | 1280 | 640 | 320 | 640 | 1280 | 640 | 2560 | 1280 | 1280 | 1280 | 1280 |
| A/Bulgaria/552/2016 |  | 6B. 1 | 2016-02-17 | SIAT2/MDCK1 | 1280 | 640 | 320 | 640 | 1280 | 640 | 2560 | 1280 | 1280 | 1280 | 1280 |
| A/Bulgaria/671/2016 |  | 6B. 1 | 2016-02-19 | SIAT2/MDCK1 | 1280 | 640 | 320 | 640 | 1280 | 640 | 2560 | 1280 | 1280 | 2560 | 1280 |
| A/Bulgaria/670/2016 |  | 6B. 1 | 2016-02-19 | SIAT2/MDCK1 | 1280 | 640 | 320 | 640 | 640 | 640 | 2560 | 1280 | 1280 | 2560 | 1280 |
| A/Bulgaria/665/2016 |  | 68. 1 | 2016-02-19 | SIAT2/MDCK1 | 1280 | 640 | 320 | 640 | 640 | 640 | 2560 | 1280 | 1280 | 1280 | 1280 |
| A/Bulgaria/694/2016 |  | 6B. 1 | 2016-02-22 | SIAT1/MDCK1 | 1280 | 640 | 320 | 640 | 1280 | 640 | 1280 | 2560 | 1280 | 2560 | 2560 |
| A/Bulgaria/691/2016 |  | 6 B .1 | 2016-02-22 | SIAT1/MDCK1 | 640 | 640 | 320 | 320 | 640 | 320 | 1280 | 1280 | 640 | 1280 | 1280 |
| A/Bulgaria/811/2016 |  | 68. 1 | 2016-03-04 | SIAT2/MDCK1 | 1280 | 640 | 320 | 640 | 1280 | 640 | 2560 | 1280 | 1280 | 2560 | 2560 |

[^4]Table 3-2. Antigenic analysis of A(H1N1)pdm09 viruses by HI


* Superscripts refer to antiserum properties ( $<$ relates to the lowest dilution of antiserum used)
$1<=<40 ;{ }^{2}<=<80$

Table 3-3. Antigenic analysis of A(H1N1)pdm09 viruses by HI

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[^5]Figure 1. Phylogenetic comparison of influenza A(H1N1)pdm09 HA genes


## Influenza A(H3N2) virus analyses

As described in many previous reports ${ }^{3}$, influenza A(H3N2) viruses continue to be difficult to characterise antigenically by HI assay due to variable agglutination of red blood cells (RBCs) from guinea pigs, turkeys and humans or the loss of the ability of viruses to agglutinate any of the RBCs. This is a particular problem for most viruses that fall in genetic subgroup 3C.2a, as was highlighted first in the November 2014 report ${ }^{4}$.
Results of HI tests performed with guinea pig RBCs in the presence of 20 nM oseltamivir, added to circumvent NAmediated binding of $\mathrm{A}(\mathrm{H} 3 \mathrm{~N} 2)$ viruses to the RBCs, are shown in Table 4. Three test viruses, from Romania, retained sufficient HA titre to be analysed by HI assay; genetic characterisation of these is ongoing.
The test viruses, propagated in MDCK-SIAT1 cells, reacted poorly in HI assays with the panel of post-infection ferret antisera relative to the titres of the antisera with their respective homologous viruses (shown in red: Table 4). However, the antisera raised against A/Stockholm/6/2014 (3C.3a: tissue culture- and egg-propagated), tissue culture-propagated A/Hong Kong/5738/2014 (3C.2a), tissue culture-propagated A/Georgia/532/2015 and A/Hong Kong/4801/2014 (3C.2a: tissue culture- and egg-propagated), gave reactivity with all test viruses. The antiserum raised against egg-propagated A/Switzerland/9715293/2013 (3C.3a), the northern hemisphere 2015-16 vaccine component, reacted with test viruses at titres reduced at least eightfold compared to the homologous titre, while antiserum raised against egg-propagated A/Hong Kong/4801/2014, the virus recommended for use in vaccines for the southern hemisphere 2016 and northern hemisphere 2016-17 influenza seasons, yielded titres reduced by two-to-fourfold compared to the homologous titre and gave absolute titres of 80 or greater.

Since 2009, seven genetic groups based on the HA gene have been defined for A(H3N2) viruses. Phylogenetic analysis of the HA genes of representative $A(H 3 N 2)$ viruses with recent collection dates is shown in Figure 2. The HA genes fall within clade 3C. This clade has three subdivisions: 3C. 1 (represented by A/Texas/50/2012, the vaccine virus recommended for use in the 2014-15 northern hemisphere season), 3C. 2 and 3C.3. Viruses in these three subdivisions had been antigenically similar. In 2014 three new subclades emerged, one in subdivision 3C.2, 3C.2a, and two in 3C.3, 3C.3a and 3C.3b, with subclade 3C. 2 a viruses dominating in recent months (Figure 2). While viruses in subclades 3C.2a and 3C.3a are antigenic drift variants, those in 3C.3b have remained antigenically similar to previously circulating viruses in the 3C. 3 subdivision. Amino acid substitutions that define these subdivisions and subclades are:

- (3C.2) N145S in HA1, and D160N in HA2, e.g. A/Hong Kong/146/2013
- (3C.2a) Those in 3C. 2 plus L3I, N144S (resulting in the loss of a potential glycosylation site), F159Y, K160T (in the majority of viruses, resulting in the gain of a potential glycosylation site), N225D and Q311H in HA1, e.g. A/Hong Kong/5738/2014
- (3C.3) T128A (resulting in the loss of a potential glycosylation site), R142G and N145S in HA1, e.g. A/Samara/73/2013
- (3C.3a) those in $3 C .3$ plus A138S, F159S and N225D in HA1, many with K326R, e.g. A/Switzerland/9715293/2013
- (3C.3b) those in 3C. 3 plus E62K, K83R, N122D (resulting in the loss of a potential glycosylation site), L157S and R261Q in HA1 with M18K in HA2, e.g. A/Netherlands/525/2014

Based on results available at the time of the February 2015 vaccine composition meeting showing cross-reactivity of antisera raised against subclade 3C.3a and 3C.2a viruses, but with changes acquired on egg-adaptation of genetic subgroup 3C. $2 a$ viruses and, at that time, the lack of a suitable 3 C . 2 a vaccine candidate, the World Health Organization recommendation was to use an A/Switzerland/9715293/2013-like virus as the A(H3N2) component of vaccines for the northern hemisphere 2015-16 influenza season [1]. After February 2015, a new subclade designated 3C.3b emerged, these three subclades being antigenically distinguishable, but subclade 3C.2a viruses became prevalent and have remained so. While ferret antisera raised against 3C.3a and 3C.2a subclade viruses showed some cross-reactivity with viruses in all three subclades, antisera raised against 3C.3b viruses were subclade specific. With the availability of new subclade 3 C . 2 a vaccine candidates and the continued crossreactivity of antisera raised against viruses in subclades 3C.3a and 3C.2a viruses, the World Health Organization recommendation for the A(H3N2) component of influenza vaccines for the southern hemisphere 2016 [2] and northern hemisphere 2016-17 [3] influenza seasons was for an A/Hong Kong/4801/2014-like (3C.2a) virus.

[^6]Table 4. Antigenic analysis of A(H3N2) viruses by HI (guinea pig RBC with 20nM oseltamivir)


Figure 2. Phylogenetic comparison of influenza A(H3N2) HA genes


## Influenza B virus analyses

EU/EEA countries have provided 134 influenza type B viruses, of which 130 were ascribed to a lineage: 114 B/Victoria-lineage and 16 B/Yamagata-lineage (Table 2).

## Influenza B - Victoria lineage

Since the April 2016 report 22 viruses of this lineage from Bulgaria and Romania have been characterised antigenically. HI results are shown in Table 5; genetic characterisation is ongoing.

The test viruses showed similar HI reactivity patterns to those from the 2014-15 influenza season: greater than eightfold reductions in HI titres compared with the titre for the homologous virus with post-infection ferret antisera raised against the recommended vaccine virus for quadrivalent live and inactivated vaccines for the northern hemisphere 2015-2016 influenza season, B/Brisbane/60/2008. Similarly, they were poorly recognised by post-infection ferret antisera raised against the reference viruses propagated in eggs $B / M a l t a / 636714 / 2011$, $\mathrm{B} /$ Johannesburg/3964/2012 and $\mathrm{B} /$ South Australia/81/2012. In contrast, all test viruses showed reactivity within fourfold, the majority within twofold, of the titres for the corresponding homologous viruses with antisera raised against viruses that are considered to be surrogate tissue culture-propagated antigens representing the eggpropagated $\mathrm{B} /$ Brisbane/60/2008 prototype virus; these antisera were raised against tissue-culture-propagated viruses B/Hong Kong/514/2009 (clade 1B), B/Ireland/3154/2015 and B/Nordrhein-Westfalen/1/2016 (both clade 1 A ).

Phylogenetic analysis of the HA gene of representative B/Victoria lineage viruses is shown in Figure 3. Throughout the previous season and this season to date, viruses from Europe and elsewhere have HA genes that fall into the $B / B r i s b a n e / 60 / 2008$ clade (clade 1 A ) and remain antigenically similar to the vaccine virus B/Brisbane/60/2008. The great majority of viruses, with collection dates since October 2015, fall in a major subcluster defined by amino acid substitutions I117V, N129D and V146I within clade 1A.

These results, linked with the rise in the proportion of B/Victoria-lineage viruses seen in the 2015 southern hemisphere and 2015-2016 northern hemisphere influenza seasons, support the recommendations made to include $\mathrm{B} /$ Brisbane/60/2008 in trivalent influenza vaccines for the southern hemisphere 2016 [2] and northern hemisphere 2016-2017 [3] influenza seasons and in the quadrivalent vaccines.

## Influenza B - Yamagata lineage

HI results for one $B /$ Yamagata-lineage test virus analysed since the April 2016 report are shown in Table 6, and genetic characterisation is ongoing.
The homologous titres of the 10 post-infection ferret antisera, shown in red, ranged from 160-640, and the test virus showed reactivity with all 10 (Table 6).

Antisera raised against egg-propagated clade 3 viruses B/Phuket/3073/2013 (the virus recommended for inclusion in trivalent influenza vaccines for the northern hemisphere 2014-2015 season) and B/Hong Kong/3417/2014 recognised the test virus at titres within twofold of their respective homologous titres. The test virus showed fourfold or greater reductions in HI reactivity, compared to homologous titres, with the other eight antisera, including that raised against egg-propagated B/Massachusetts/02/2012, the clade 2 vaccine virus recommended for use in the 2014-15 northern hemisphere influenza season.
Figure 4 shows a phylogenetic analysis of the HA genes of representative B/Yamagata-lineage viruses. Worldwide, the vast majority of HA genes from recently collected viruses have fallen in the B/Wisconsin/1/2010B/Phuket/3073/2013 clade (clade 3), with the great majority falling in a subgroup defined by HA1 L172Q amino acid substitution. A few viruses, annotated in the phylogenetic tree (e.g. B/Oman/6341/2015), are reassortants carrying NA genes normally associated with the B/Victoria-lineage.
Based on such results, a B/Phuket/3073/2013-like virus has been recommended for inclusion in quadrivalent vaccines for the 2016 southern hemisphere [2] and 2016-2017 northern hemisphere [3] influenza seasons.

Table 5. Antigenic analysis of influenza B/Victoria-lineage viruses by HI


Figure 3. Phylogenetic comparison of influenza B/Victoria-lineage HA genes


Table 6. Antigenic analysis of influenza B/Yamagata-lineage viruses by HI


Figure 4. Phylogenetic comparison of influenza B/Yamagata-lineage HA genes


## Summary of genetic data submitted to TESSy

For the period covering weeks 40/2015-20/2016, 2601 viruses have been characterised genetically: 1770 A(H1N1)pdm09 clade 6B represented by A/South Africa/3626/2013 (6B.1 and 6B.2 subclade designations were not available as reporting categories at the start of the 2015-2016 influenza season); $211 \mathrm{~A}(\mathrm{H} 3 \mathrm{~N} 2)$ subclade 3 C .2 a represented by A/Hong Kong/4801/2014, 65 subclade 3C.3a represented by A/Switzerland/9715293/2013, two subclade 3C.3b represented by A/Stockholm/28/2014, and two subclade 3C. 3 represented by A/Samara/73/2013; 496 B/Victoria-lineage clade 1A represented by B/Brisbane/60/2008; and 55 B/Yamagata-lineage clade 3 represented by B/Phuket/3073/2013.
Note: the numbers quoted above are reduced compared to the March and April reports as a consequence of correcting a problem encountered with the reporting of virus characterisation data to TESSy.

## Antiviral susceptibility

For weeks 40/2015-20/2016 of the 2015-2016 influenza season, countries reported on the antiviral susceptibility of 2700 A(H1N1)pdm09 viruses, 172 A(H3N2) viruses and 523 influenza type B viruses from sentinel and nonsentinel sources. All but 31 showed no molecular or phenotypic evidence of reduced inhibition (RI) by neuraminidase inhibitors (oseltamivir and zanamivir). Twenty-six A(H1N1)pdm09 viruses carried NA H275Y amino acid substitution associated with highly reduced inhibition (HRI) by oseltamivir, one A(H3N2) virus showed RI by oseltamivir associated with NA-E119V amino acid substitution and four B/Victoria-lineage viruses showed HRI by both drugs due to NA-R374K amino acid substitution.
Phenotypic testing for susceptibility to oseltamivir and zanamivir has been conducted on 501 viruses at the WIC: 316 A(H1N1)pdm09, 69 A(H3N2), 99 B/Victoria-lineage and 17 B/Yamagata-lineage viruses. All but one A(H1N1)pdm09 virus showed normal inhibition (NI) by these neuraminidase inhibitors: A/Bayern/151/2015 showed reduced inhibition (RI) by zanamivir and carried NA I117R amino acid substitution.

## Influenza A(H7N9) virus

On 1 April 2013, the World Health Organization (WHO) Global Alert and Response [4] reported that the China Health and Family Planning Commission notified the WHO of three cases of human infection with influenza A(H7N9). The cases were confirmed by laboratory testing on 29 March 2013 by the Chinese CDC. A description of the characteristics of H7N9 viruses can be found on the WHO website [5]. Increased numbers of cases were reported over the course of the 2013-14, 2014-15 and 2015-16 seasons and cases have been reported recently [6]. A revised Rapid Risk Assessment [7] for these A(H7N9) viruses was carried out by ECDC and posted on 2 February 2015. WHO posted a summary of human infection on 31 January 2014 [8], updated on 09 May 2016 [9] with 18 new cases since the report of 04 April 2016, and conducted a risk assessment on 23 February 2015 [10]. In light of the assessment, WHO advised that countries continue to strengthen influenza surveillance. WHO last summarised the numbers of cases of human infection related to their geographic location on 14 July 2014 [11] and has provided subsequent situation updates, with the latest dated 17 May 2016 [6].

## Influenza A(H5) virus

The most recent monthly risk assessment of influenza at the human-animal interface was published by WHO on 9 May 2016 [9]. Since the last WHO Influenza update on 4 April 2016, three laboratory-confirmed human cases of avian influenza A(H5N6) virus infection in China have been reported to WHO. While no human cases of A(H5N1) infection in Egypt have been reported for the same period, a single case of human infection with A(H9N2) was reported. ECDC published an updated rapid risk assessment on the situation in Egypt on 13 March 2015 [12] and an epidemiological update 10 April 2015 [13]. On 2 December 2015, ECDC published a rapid risk assessment related to identification highly pathogenic H 5 viruses in poultry in France [14].

## WHO CC reports

A description of results generated by the WHO Collaborating Centre for Reference and Research on Influenza at the Crick Worldwide Influenza Centre, the Francis Crick Institute, Mill Hill Laboratory and used at the WHO Vaccine Composition Meetings held in Memphis, USA, 21-23 September 2015, and at WHO Geneva, 22-24 February 2016, can be found at:

## Note on the Figures

The phylogenetic trees were constructed using RAxML, drawn using FigTree and annotated using Adobe Illustrator. The bars indicate the proportion of nucleotide changes between sequences. Reference strains are viruses to which post-infection ferret antisera have been raised. The colours indicate the month of sample collection. Isolates from WHO NICs in EU/EEA countries are marked (\#). Sequences for some viruses from non-EU/EEA countries were recovered from GISAID. We gratefully acknowledge the authors, originating and submitting laboratories of the sequences from GISAID's EpiFlu database which were downloaded for use in the preparation of this report (all submitters of data may be contacted directly via the GISAID website), along with all laboratories who submitted sequences directly to the London WHO Collaborating Centre.

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[^0]:    This report was prepared by Rod Daniels, Vicki Gregory, Burcu Ermetal, Aine Rattigan and John McCauley on behalf of the European Reference Laboratory Network for Human Influenza (ERLI-Net), under contract to the European Centre for Disease Prevention and Control (ECDC).
    Suggested citation: European Centre for Disease Prevention and Control. Influenza virus characterisation, summary Europe, May 2016. Stockholm: ECDC; 2016.
    (C) European Centre for Disease Prevention and Control, Stockholm, 2016.

    Reproduction is authorised, provided the source is acknowledged.

[^1]:    ${ }^{\text {a }}$ There was a data problem in TESSy relating to weeks 16-19/2016 with double entry of data from some countries. It affected sentinel, non/sentinel detections and SARI data. This problem was identified in week 19 and corrected. As a consequence the data included in the April report (to week $17 / 2016$ ) was incorrect - the numbers given above (to week 20/2016) are correct, but reduced compared to the same table in the April report.

[^2]:    ${ }^{1}$ European Centre for Disease Prevention and Control. Influenza virus characterisation, summary Europe, April 2016. Stockholm: ECDC; 2016. Available from: http://ecdc.europa.eu/en/publications/Publications/ERLI-Net\%20report\%20April\%202016.pdf

[^3]:    ${ }^{2}$ European Centre for Disease Prevention and Control. Influenza virus characterisation, summary Europe, September 2014. Stockholm: ECDC; 2014. Available from: http://ecdc.europa.eu/en/publications/Publications/Influenza-ERLI-Net-report-Sept2014.pdf

[^4]:    * Superscripts refer to antiserum properties (< relates to the lowest dilution of antiserum used)

[^5]:    * Superscripts refer to antiserum properties (< relates to the lowest dilution of antiserum used)

[^6]:    ${ }^{3}$ For example, the September 2013 report: European Centre for Disease Prevention and Control. Influenza virus characterisation, summary Europe, September 2013. Stockholm: ECDC; 2013. Available from:
    http://www.ecdc.europa.eu/en/publications/Publications/influenza-virus-characterisation-sep-2013.pdf
    ${ }^{4}$ European Centre for Disease Prevention and Control. Influenza virus characterisation, summary Europe, November 2014. Stockholm: ECDC; 2014. Available from: http://www.ecdc.europa.eu/en/publications/Publications/ERLI-Net report November 2014.pdf

